

### **Lead Inventors:**

### Dan Landau, M.D., Ph.D.

Associate Professor of Medicine, Division of Hematology and Medical Oncology Associate Professor of Physiology and Biophysics, Weill Comell Medicine Core Member, New York Genome Center

### **Business Development Contact:**

Jamie Brisbois
Manager, Business Development and Licensing

(646) 962-7049 jamie.brisbois@cornell.edu

### Background & Unmet Need

- Over 1,600 transcription factors orchestrate gene regulation through complex DNA interactions, which are often disrupted in disease and aging
- Current bulk methods like ChIP-seq and CUT&TAG require stringent conditions that disrupt weaker protein-DNA interactions and mask cellular heterogeneity by providing only population averages
- While techniques like DamID can map protein-DNA interactions genome-wide, they cannot capture the dynamic regulatory networks at single-cell resolution needed to understand cell-type specific responses and disease mechanisms
- Unmet Need: Methods that can directly identify and map non-histone protein binding events with singlecell resolution

### **Technology Overview**

- The Technology: A sequencing method (DnD-seq) that combines nanobody-directed targeting with controlled DNA deamination to record protein-DNA binding events at single-cell resolution
- The technology utilizes an engineered split DddA enzyme with nanobody targeting enables controlled, protein-specific DNA modification that permanently marks binding sites
- The system remains inactive until deliberately activated, ensuring specific recording of protein-DNA interactions
- PoC Data: Demonstrated high specificity with clear identification of transcription factor footprints and concordance with ChIP-seq reference data
- Demonstrated CTCF profiling in primary human CD8 T cells, revealing how IDH2 mutations alter CTCF binding patterns and chromatin organization across different cell types, validated by genotype-specific analysis

### Inventors:

Dan Landau Ivan Raimondi Wei-Yu Chi

### Patents:

Provisional Filed

### **Publications:**

Chi et al. bioRxiv [Preprint]. 2025

### **Biz Dev Contact:**

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### Cornell Reference:

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DddA: Double-stranded DNA deaminase A CTCF: CCCTC-binding factor I DH2: Isocitrate dehydrogenase 2

### **Technology Applications**

- Enhanced AI/ML cell modeling through highresolution protein-DNA interaction data
- Optimization of chromatin-targeted therapeutic development
- Improved detection of off-target effects in genetic engineering
- Advanced cellular reprogramming through precise transcription factor mapping

### **Technology Advantages**

- Integration with diverse platforms (PTA, DLP, 10X Multiome) for comprehensive analysis
- Enables simultaneous profiling of protein binding and chromatin accessibility at single-cell resolution
- Reveals cellular heterogeneity previously masked in bulk approaches
- Expandable platform allowing incorporation of additional protein-specific nanobodies

# DddA full length 108aa DddA\_NT (inactīve) Supporting Data / Figures 25aa DddA\_NT (activator)

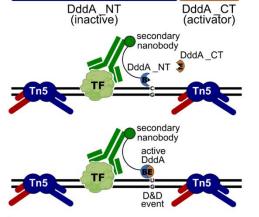


Figure 1: Schematic representation of the DnD-seq split enzyme system. The engineered construct comprises an inactive DddA N-terminal domain fused to a nanobody that recognizes target antibodies, and a separate C-terminal domain. When brought together at antibody-bound transcription factor (TF) sites, the enzyme becomes active.

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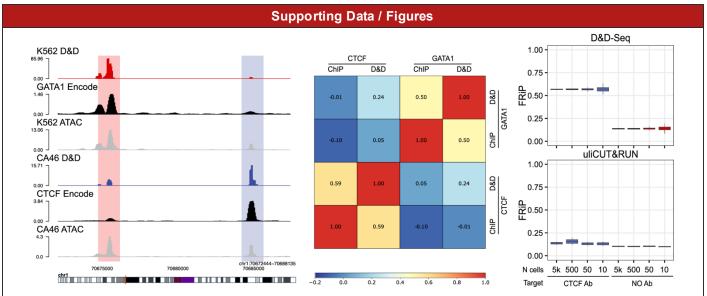


Figure 2: Left: DNA binding locations shown as peaks - red peaks show where GATA1 binds in K562 cells, blue peaks where CTCF binds in CA46 cells, with black lines showing reference data.

Middle: Correlation matrix demonstrating strong matches between D&D-seq and reference data for each protein.

Right: Comparison using Fraction of Reads in Peaks (FRIP) shows D&D-seq achieves 57% of DNA reads in known binding sites versus only 14% for current methods, maintaining this performance even with as few as 10 cells.

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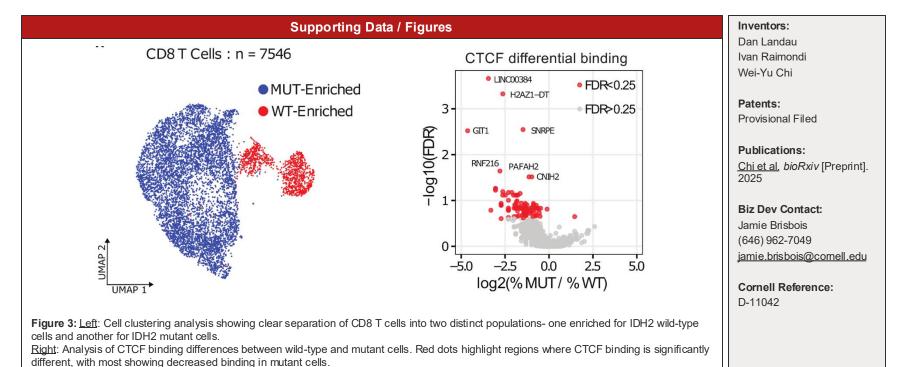
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# DnD-Seq: Simultaneous Profiling of Transcription Factor Occupancy and Accessible Chromatin





## Weill Cornell Medicine